Degradation of Salicylate, an Important Plant-Signaling Molecule, by the Fungal Plant Pathogen Sclerotinia sclerotiorum



Cory D. Penn and Steven L. Daniel Department of Biological Sciences, Eastern Illinois University, Charleston, Illinois



Introduction

Sclerotinia sclerotiorum is a nectrotrophic phytopathogen (Figure 1) that has the ability to infect up to 406 plant species, including many cash crops [6]. The plant pathogen is capable of significantly reducing crop Agar (PDA) plates at 25°C and transferred (5 mm plug) to yields as it has been estimated that annually it accounts for \$200 billion 50 mL of culture medium. Culture medium (pH 6.5) losses in the United States alone [2]. The key pathogenic trait of S. sclerotiorum is its ability to synthesize oxalate (toxin) [1]. It has been 0.1% soytone, 50 mM MES (buffer), glucose, and proposed that isolates of *S. sclerotiorum* that produce more oxalate tend to be more pathogenic than strains that produce less [4].

One circumstance that has been shown to regulate oxalate cultures were pulled from the incubator and harvested. production by the organism is the type of carbon sources available in the environment [4]. External pH and buffering capacity of the environment harvested by the use of a vacuum system and captured also plays a large role in oxalate production by *S. sclerotiorum* [3]. It has by Whatman® 70 mm filter paper. The biomass was then been suggested from past studies that increasing the culture pH or dried at 50°C for 72 hours, cooled in a desiccator, and buffering capacity of the medium, increases the ability of the organism to then weighed with the use of an electronic scale. produce oxalate [3].

Salicylate is a plant-signaling molecule synthesized by plants in separated from growth by use of filter paper. The pH of response to a pathogen attack and is involved in activating plant defenses. The compound plays a role in systematic acquired resistance (SAR), 230A pH meter and an Orion semi-micro combination which is the process by which older, infected leaves causes the development of resistance in younger leaves [6]. Very little work has been 🛮 by 🛮 a 🔻 Beckman Gold high performance liquid done to determine whether S. sclerotiorum has the ability to metabolize salicylate, and if it does, what effect it has on the metabolism and growth of the organism.

Goals

- To determine whether salicylate was degraded by S. sclerotiorum
- To determine the impact of salicylate on biomass formation, oxalate production, and culture acidification by S. sclerotiorum

Methods and Materials

Growth of Culture. S. sclerotiorum isolates (D-E7, 105, and W-15HT) were maintained on Potato Dextrose consisted of minerals and trace metals supplemented by salicylate. Cultures were incubated for 7 days while shaking at 200 rpm. After the 7 days of incubation the

Biomass (Growth) Determination. Biomass was

Substrate Level Determination. Filtrate was the filtrate was determined with the use of an Orion model electrode. A 1 mL sample was also collected for analysis chromatograph fitted with a 300 mm Bio-Rad Aminex HPX-87H column. Oxalate and salicylate detections were done at 210 nm and the glucose was detected by the refractive index detector. All concentrations of the compounds were expressed on the millimolar (mM) basis. Throughout the study, no distinctions were made between organic acids and their salt forms.

Results

- Salicylate was degraded by all three isolates of S. sclerotiorum; however, salicylate was not supportive (Table 1).
- Uninoculated controls showed that salicylate was not abiotically degraded under culture conditions (data not shown). Also, salicylate was not synthesized by S. sclerotiorum during growth in the medium (Table 1).
- Salicylate was not growth supportive and at higher concentration (10 mM) seemed to inhibit growth (Table 1).

Table 1. Salicylate degradation and biomass accumulation by *S. sclerotiorum* isolates D-E7, 105, and W-15HT. Salicylate Degradation (mM) Biomass Produced (a)

		Salicylate Degradation (mivi)			Biomass Produced (g)			
	Addition	D-E7	105	W-15HT	D-E7	105	W-15HT	
	None	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	0.020 ± 0.002	0.009 ± 0.001	0.017 ± 0.001	
7	1 mM Salicylate	0.24 ± 0.05	0.75 ± 0.05	0.00 ± 0.00	0.022 ± 0.004	0.006 ± 0.003	0.015 ± 0.002	
	10 mM Salicylate	6.05 ± 1.62	9.04 ± 0.27	3.14 ± 0.98	0.029 ± 0.004	0.012 ± 0.002	0.025 ± 0.004	
	25 mM Glucose		0.00 ± 0.00		0.137 ± 0.002	0.061 ± 0.028	0.138 ± 0.068	
	25 mM Glucose + 1 mM Salicylate	0.18 ± 0.30	0.00 ± 0.00	0.00 ± 0.00	0.125 ± 0.007	0.057 ± 0.003	0.117 ± 0.018	
	25 mM Glucose + 10 mM Salicylate	2.95 ± 3.90	7.09 ± 1.05	2.50 ± 3.76	0.087 ± 0.002	0.030 ± 0.003	0.066 ± 0.025	



Literature Cited

- 1. Boland GJ, Hall R (1994) Index of plant hosts of Sclerotinia sclerotiorum. Can J Plant Pathol 16:93-108.
- 2. Bolton MD, Thomma BPHJ, Nelson BD (2006) Sclerotinia sclerotiorum (Lib.) de Bary: biology and molecular traits of a cosmopolitan pathogen. Mol Plant Pathol 7:1-16.
- 3. Culbertson BJ, Krone J, Gatebe E, Furumo NC, Daniel SL (2007) Impact of carbon sources on growth and oxalate synthesis by the phytopathogenic fungus Sclerotinia Sclerotinia Sclerotiorum. World J Microbiol Biotechnol 23:1357-1362. Culbertson BJ, Furumo NC, Daniel SL (2007) Impact of nutritional supplements and monosaccharides on growth, oxalate accumulation, and culture pH by Sclerotinia sclerotiorum. FEMS Microbiol Lett 270:132-138. Karegoudar TB, Kim CK (200) Microbial Degradation of Monohydroxybenzoic Acids. Journal of Microbiology 38: 53-61.
- 6. Stotz, X. G. a. H. U. (2007). "Defense Against Sclerotinia sclerotiorum in Arabidopsis is Dependent on Jasmonic Acid, Salicylic Acid, and Ethylene Signaling." Molecular Plant-Microbe Interactions 20(11):1384-1395

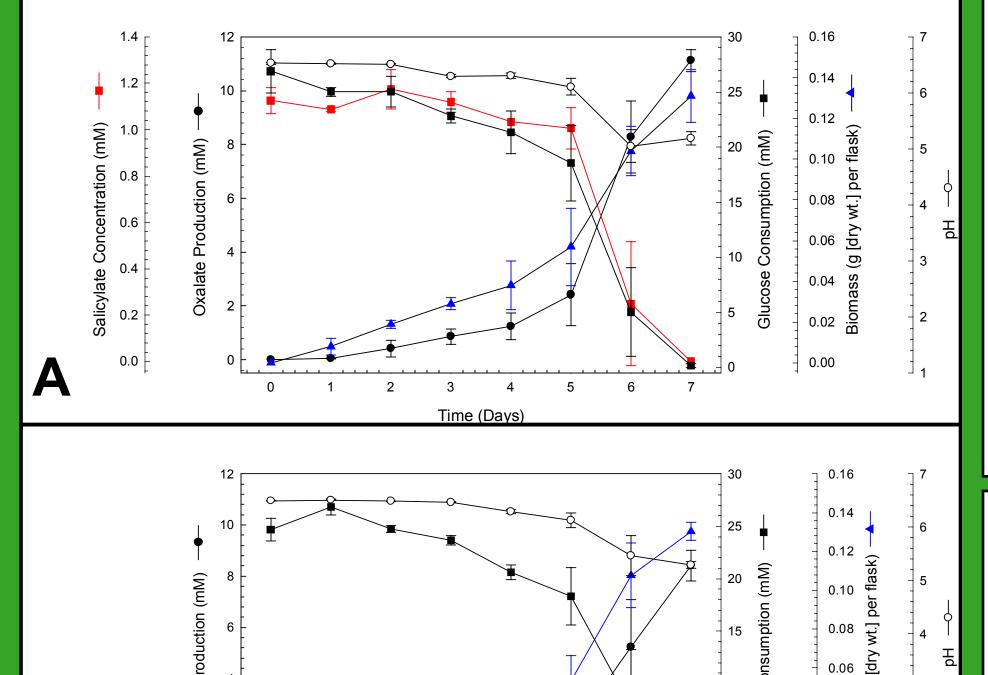
- Salicylate did not enhance nor repress oxalate production in the presence of glucose (Table 2).
- Salicylate did not enhance nor repress the acidification of the medium (Table 2).

Table 2. Effect of salicylate degradation on biomass and oxalate produced, glucose consumption, and acidification of the medium by S. sclerotiorum D-E7.

Addition	Salicylate (mM)	Biomass (g)	Oxalate (mM)	Glucose (mM)	Final pH
None	0.00 ± 0.00	0.020 ± 0.002	3.24 ± 0.22	0.00 ± 0.00	6.46 ± 0.02
1 mM Salicylate	0.24 ± 0.05	0.022 ± 0.004	4.33 ± 0.18	0.00 ± 0.00	6.39 ± 0.02
10 mM Salicylate	6.05 ± 1.62	0.029 ± 0.004	7.83 ± 1.33	0.00 ± 0.00	6.24 ± 0.07
25 mM Glucose	0.00 ± 0.00	0.137 ± 0.002	11.45 ± 0.18	0.00 ± 0.00	5.26 ± 0.04
25 mM Glucose + 1 mM Salicylate	0.18 ± 0.30	0.125 ± 0.007	9.81 ± 2.17	1.29 ± 2.23	5.24 ± 0.11
25 mM Glucose + 10 mM Salicylate	2.95 ± 3.90	0.087 ± 0.002	12.84 ± 2.50	1.58 ± 2.74	4.92 ± 0.21

0.04

0.02



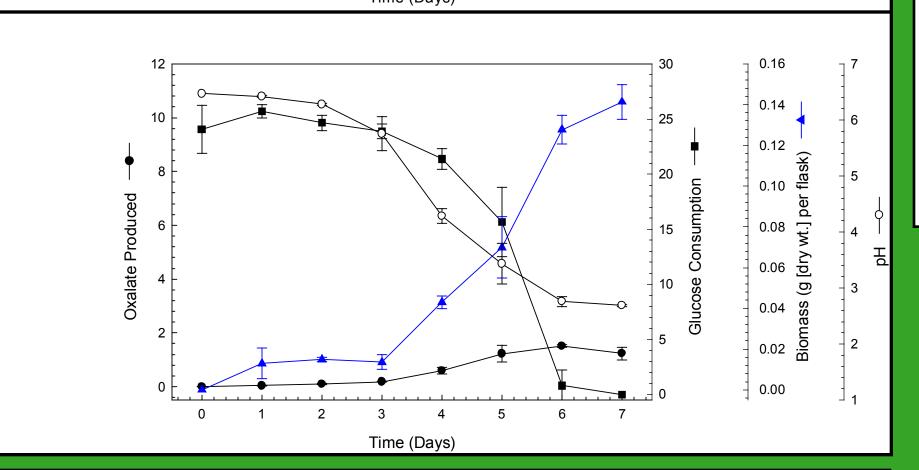


Figure 2. Time course analysis of growth, oxalate accumulation, glucose and salicylate consumption, and culture pH for S. sclerotiorum D-E7 growth in medium containing 0.1% soytone, 50 mM MES, 25 mM glucose, and 1 mM salicylate (A); 0.1% soytone, 50 mM MES, and 25 mM glucose (B); and 0.1% soytone and 25 mM glucose (C).

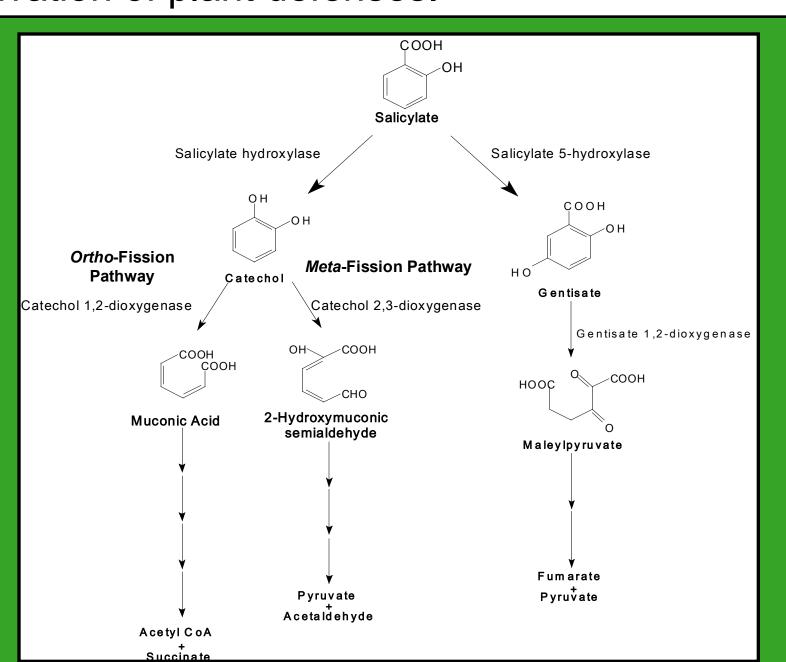
Acknowledgements

Funding for this project was provided through 2 Biological Sciences undergraduate research grants (2008), a College of Sciences seed grant (2009), and PIF grant (2008), all from Eastern Illinois University. Cultures of S. sclerotiorum isolates were obtained from the National Soybean Pathogen Collection Center at the University of Illinois in Champaign-Urbana.

- Salicylate in the culture medium does not repress growth or oxalate synthesis of *S. sclerotiorum* (Figure 2).
- All glucose and salicylate is consumed during time course, and each correlate positively which the growth of the organism and the production of oxalate (Figure 2).
- From HPLC analysis of the time course study, potential intermediates from the degradation of salicylate were observed (data not shown).

Summary

Many microorganisms have the ability to degrade salicylate, including Streptomyces, Pseudomonas, and *Micrococcus* species [6]. It is still unknown whether this same pathway (**Figure 3**) is followed by S. sclerotiorum. However, the ability of S. sclerotiorum to degrade salicylate may allow this phytopathogen to shut down plant defenses by intercepting degrading plant-signaling molecules that are directly linked with the activation of plant defenses.



Possible pathway of salicylate degradation by Sclerotinia sclerotiorum that is observed in other microorganisms [6].