

Oxalate Consumption by Probiotic Microorganisms

I-048

Steven L. Daniel and Ryan Cox

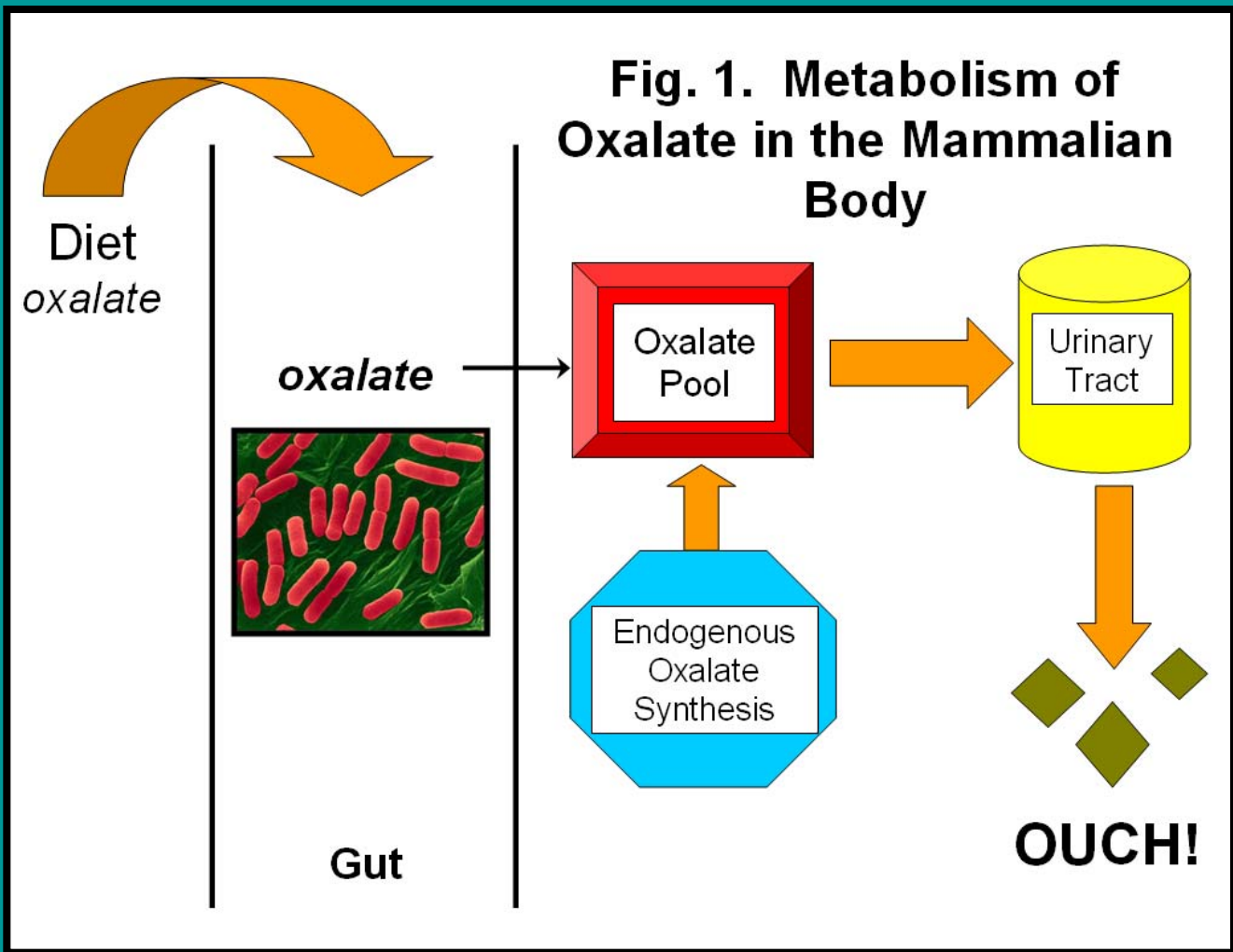
Department of Biological Sciences, Eastern Illinois University, Charleston, IL



Introduction

Oxalate and Kidney Stones

- Oxalate is found in human and animal diets
- Oxalate is also produced endogenously as an end product of cellular metabolism
- Oxalate (absorbed from diet and endogenously synthesized) eliminated by urinary excretion
- High levels of oxalate in the urine (hyperoxaluria) can lead to kidney stones (see Figure 1)



Risk Factors for Kidney Stones

- Increased dietary oxalate intake
- Increased endogenous oxalate synthesis
- Increased absorption of oxalate due to the absence of oxalate-degrading bacteria in the gut
- Oxalate-degrading gut bacteria impact oxalate metabolism in humans and animals**
- Oxalate-degrading gut bacteria represent novel therapies for the prevention and treatment of kidney stones**

Oxalobacter formigenes

- Primary oxalate-degrading bacterium in the intestinal tracts of humans and animals (Allison et al., 1995)
- Anaerobe (a "specialist"): uses only oxalate as an energy source and converts it to formate and CO₂
- Colonization starts at 1 year of age in humans
- 70-80% of adult population are colonized
- Absence from the gut has been linked to increased urinary oxalate excretion and increased risk of kidney stone formation
- Currently being developed as a medical probiotic to prevent hyperoxaluria and stone formation

So, are there other oxalate-degrading bacteria in the gut?

Oxalate Degradation by Gut and Probiotic Bacteria

- Eubacterium lentum* (Ito et al., 1996)
- Enterococcus faecalis* (Hokama et al., 2000)
- Lactic Acid Bacteria (LAB)
 - Lactobacillus acidophilus*, *Streptococcus thermophilus*, *Bifidobacterium infantis* (Campieri et al., 2001)
 - Leuconostoc lactis*, *Lactobacillus plantarum*, *Lactobacillus acidophilus* (Veeze et al., 2004)
 - Bifidobacterium lactis*, *Bifidobacterium breve*, *Bifidobacterium animalis*, *Bifidobacterium longum*, *Bifidobacterium infantis*, *Bifidobacterium adolescentis* (Federici et al., 2004)
 - Lactobacillus casei* (Kwak et al., 2006)
- All the above are "generalists" (substrates, other than oxalate, used for growth during oxalate degradation)

Impact of Probiotic Bacteria on Urinary Oxalate Excretion

- Pilot study by Campieri et al. (2001)
 - Freeze-dried mix of 5 Lactic Acid Bacteria (LAB) (10¹¹/g, *Lactobacillus acidophilus*, *Streptococcus thermophilus*, *Bifidobacterium infantis*, *Lactobacillus plantarum*, *Lactobacillus breve*)
 - Urinary oxalate excretion reduced in all LAB-fed patients with stone disease and mild hyperoxaluria
 - LAB mixture developed into commercial probiotic called "Oxadrop" (sold by VSL Pharmaceuticals)
- 2nd study by Lieske et al. (2005)
 - Urinary oxalate excretion reduced in Oxadrop-fed patients with inflammatory bowel disease

Objectives

- To screen commercially available human and animal probiotics for their ability to consume oxalate *in vitro*
- To evaluate the impact of commercial probiotics on oxalate degradation by *Oxalobacter formigenes* under aerobic and anaerobic conditions

Methods

Probiotics Screened

Probiotic	Manufacturer	Probiotic Microbes Present
VSL#3	VSL Pharmaceuticals	<i>S. thermophilus</i> , <i>B. breve</i> , <i>B. longum</i> , <i>B. infantis</i> , <i>L. acidophilus</i> , <i>L. plantarum</i> , <i>L. casei</i> , <i>L. bulgaricus</i>
4 x 6 Acidophilus	NOW Foods	<i>L. acidophilus</i> , <i>B. bifidum</i> , <i>B. longum</i> , <i>S. thermophilus</i> , <i>L. bulgaricus</i> , <i>L. paracasei</i>
<i>Saccharomyces boulardii</i>	Jarrow Formulas	<i>S. boulardii</i>
Jarro-Dophilus	Jarrow Formulas	<i>L. rhamnosus</i> , <i>L. casei</i> , <i>L. plantarum</i> , <i>L. acidophilus</i> , <i>B. longum</i> , <i>B. breve</i>
Kyo-Dophilus	Wakunaga of America Co.	<i>L. acidophilus</i> , <i>B. bifidum</i> , <i>B. longum</i>
Super Potent Acidophilus	Rexall	<i>L. acidophilus</i>
Acidophilus Plus	Nature's Valley	<i>L. acidophilus</i> , <i>B. bifidum</i> , <i>B. longum</i>
Fastrack	Conklin	<i>E. faecium</i> , <i>L. acidophilus</i> , <i>S. cerevisiae</i>
Pet Inoculant	Wysong	<i>B. bifidum</i> , <i>L. lactis</i> , <i>E. faecium</i> , <i>L. acidophilus</i>
<i>Oxalobacter formigenes</i>	Dr. Milt Allison	<i>O. formigenes</i> – active broth culture

Processing Probiotics and Culture Conditions

- Commercial probiotic (recommended daily dose) added to 25 ml of culture medium in 60-ml serum bottles
 - Anaerobic medium: 10 mM oxalate, 50 mM glucose, 0.1% yeast extract, minerals, metals, cysteine, resazurin, and CO₂/HCO₃⁻ in crimp-sealed bottles
 - Aerobic medium: same as above except medium lacked cysteine, resazurin, and CO₂/HCO₃⁻ and was prepared aerobically in capped bottles
- Mixed by shaking for 1 hour to resuspend probiotic
- Probiotic cultures incubated at 37°C for 48 hours
- Oxalate measured at T₀ and at T₄₈ by HPLC analysis to determine the amount of oxalate consumed

Results

- In the absence of glucose, Jarro-Dophilus, Acidophilus Plus, Super Potent Acidophilus, Kyo-Dophilus, 4x6 Acidophilus, and Fastrack consumed very little of the oxalate (0-1%) (see Table 1).
- S. boulardii* and Pet Inoculant cultures consumed 5 and 8% of the oxalate, respectively. In contrast, VSL#3 cultures consumed all (100%) of the oxalate.
- In the presence of glucose, oxalate consumption increased slightly for several of the probiotics tested (4x6 Acidophilus, *S. boulardii*, Jarro-Dophilus, Kyo-Dophilus, and Super Potent Acidophilus).
- VSL#3 cultures consumed oxalate under both anaerobic and aerobic conditions. *S. boulardii*, Jarro-Dophilus, and *O. formigenes* were more active under anaerobic conditions (see Table 2).

Table 1. Anaerobic Oxalate Consumption by 10 Different Probiotics in the Absence and Presence of Glucose

Probiotic	Dose (inoculum)	% Oxalate Consumed ^a	
		Anaerobic (- Glucose)	Anaerobic (+ Glucose)
VSL#3	5 g	100	92
4 x 6 Acidophilus	1 g	1	16
<i>Saccharomyces boulardii</i>	1 g	5	7
Jarro-Dophilus	1 g	0	4
Kyo-Dophilus	1 g	0	2
Super-Potent Acidophilus	1 g	0	1
Acidophilus Plus	0.5 g	0	0
Pet Inoculant	1 g	8	6
Fastrack	2 ml	0	0
<i>Oxalobacter formigenes</i>	0.5 ml	100	100

^a Initial oxalate concentration was 10 mM; values are the means of duplicate cultures after 48 hours of incubation.

Table 2. Oxalate Consumption by 4 Different Probiotics under Aerobic and Anaerobic Conditions

Probiotic	Dose (inoculum)	% Oxalate Consumed ^a	
		Aerobic (+ glucose)	Anaerobic (+ glucose)
VSL#3	5 g	90	94
<i>Saccharomyces boulardii</i>	1 g	0	7
Jarro-Dophilus	1 g	1	4
<i>Oxalobacter formigenes</i>	0.5 ml	0	100

^a Initial oxalate concentration was 10 mM; values are the means of duplicate cultures after 48 hours of incubation.

- Oxalate consumption by *O. formigenes* under aerobic conditions was not observed (see Figure 2; Table 2).
- Oxalate consumption was NOT observed in the *O. formigenes* + *S. boulardii* mixture under aerobic conditions indicating that the probiotic yeast failed to protect *O. formigenes*.
- Aerobic oxalate consumption was observed in the *O. formigenes* + VSL#3 mixture, although it is unclear as to whether this was due to VSL#3, to *O. formigenes* or to both (see Figure 3).
- Under anaerobic conditions, *O. formigenes* + VSL#3 mixture consumed oxalate at a much slower rate than *O. formigenes* alone, suggesting VSL#3 slightly repressed oxalate consumption by *O. formigenes*.
- Nonetheless, *O. formigenes* appeared to be very compatible with the 2 probiotics tested.

References

Allison, M. J., S. L. Daniel, and N. Cornick. 1995. Oxalate-degrading bacteria. In: S. R. Khan (ed.), Calcium oxalate in biological systems, p. 131-168. CRC Press, Inc., Boca Raton, FL.

Campieri, C., M. Campieri, V. Bertuzzi, E. Sweeney, D. Matteuzzi, S. Stafforini, F. Pinovano, C. Cereti, S. Ulisse, O. Famularo, and C. De Simone. 2001. Reduction of oxaluria after an oral course of lactic acid bacteria at high concentration. *Kidney Int.* 60:1097-1105.

Federici, F., B. Vial, R. Goh, M. F. Pasca, S. Goh, A. B. Pres, and P. Bridgi. 2004. Characterization and heterologous expression of the oxalate coenzyme A decarboxylase gene from *Bifidobacterium lactis*. *Appl. Environ. Microbiol.* 70:5068-5073.

Hojima, S., Y. Horita, C. Toma, and Y. Ogawa. 2000. Oxalate-degrading *Enterococcus faecalis*. *Microbiol. Immunol.* 44:235-240.

Ita, H., N. Miura, M. Masai, K. Yamamoto, and T. Hara. 1999. Reduction of oxalate content of food by the oxalate-degrading bacterium, *Eubacterium aerophilum* WYH-1. *Int. J. Urol.* 3:331-34.

Iwak, C., B. C. Jeong, J. H. Kim, J. J. Lee, C. S. Han, Y. J. Baek, and S. E. Lee. 2006. Prevention of nephrolithiasis by *Lactobacillus* in stone-forming rats: a preliminary study. *Urol. Res. Open First* (DOI 10.1007/s00405-006-0064-4).

Lieske, J. C., D. S. Goffman, C. De Simone, and C. Riegler. 2005. Use of a probiotic to decrease enteric hyperoxaluria. *Kidney Int.* 68:1344-1349.

Veeze, J. S., J. H. Veeze, L. Turck, and J. Roussier. 2004. Oxalate degradation by intestinal lactic acid bacteria in dogs and cats. *Vet. Microbiol.* 101:101-106.

Kidney stone images used with permission: L. C. Herring & Co. Laboratory (www.herringlab.com)

Figure 2. Oxalate Consumption by *Oxalobacter formigenes* and *Saccharomyces boulardii* under Aerobic and Anaerobic Conditions

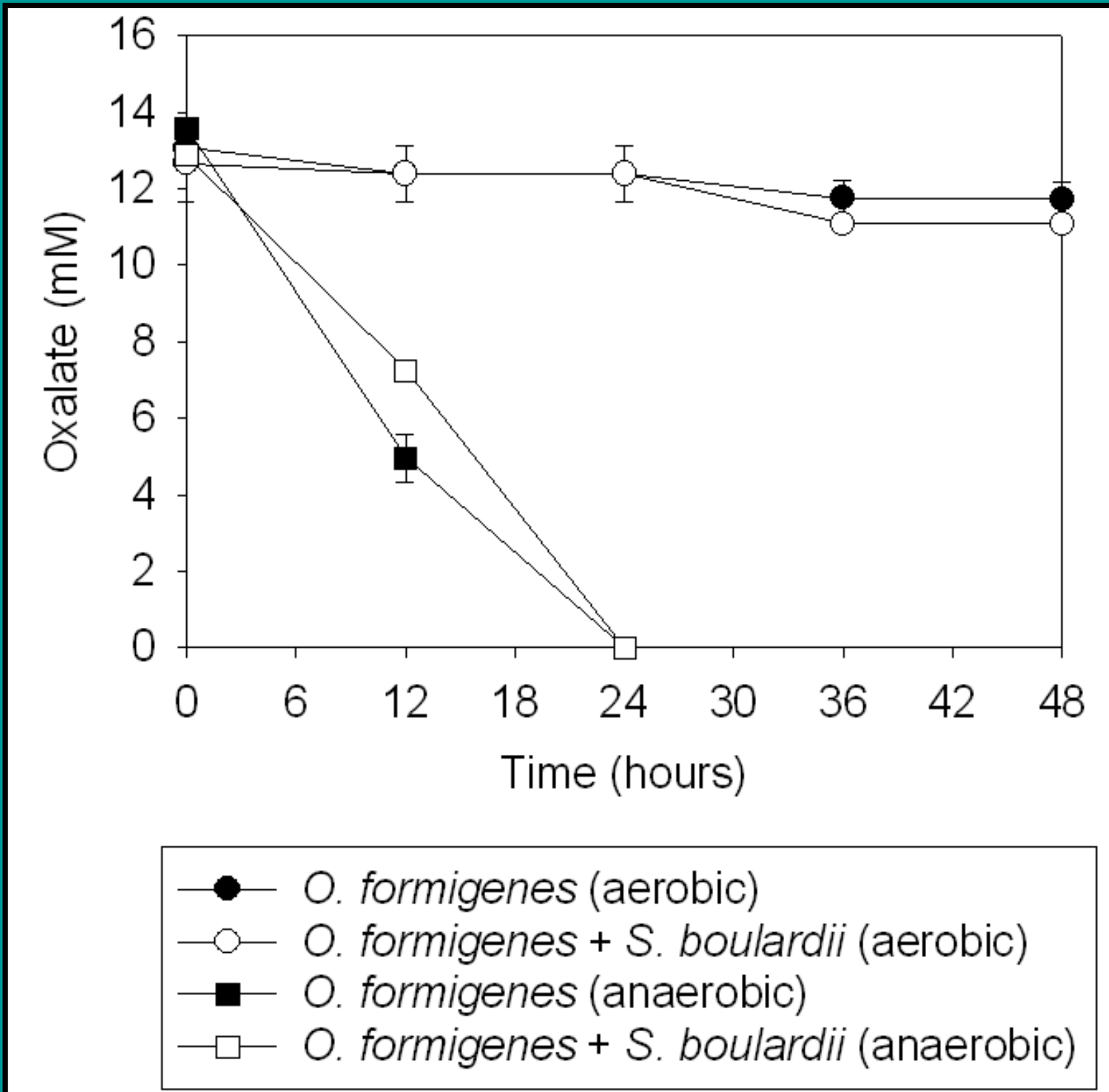
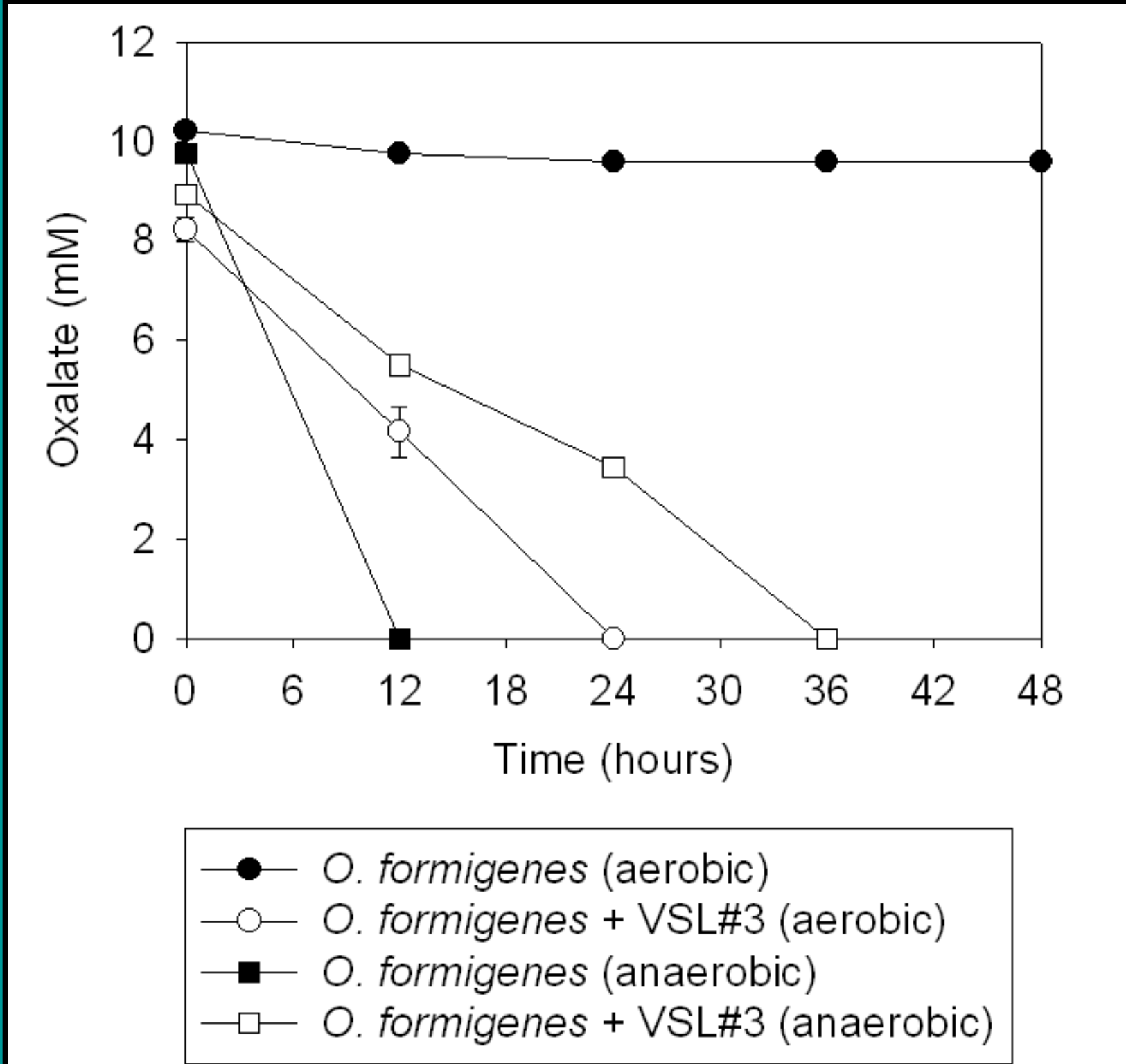


Figure 3. Oxalate Consumption by *Oxalobacter formigenes* and VSL#3 under Aerobic and Anaerobic Conditions



Summary

- Of the 9 different commercially available probiotics screened, only VSL#3 consumed significant amounts of oxalate
- Nature of the oxalate-consuming organisms or activities in VSL#3 is presently unknown
- Two probiotic (VSL#3 and *S. boulardii*) did not protect *O. formigenes* under aerobic conditions
- The combination of VSL#3 and *O. formigenes* in a single probiotic might provide a very potent and novel therapy for the prevention of calcium oxalate stones in humans and animals